Histological effect of aqueous extract of *Anacardium occidentale* (Cashew) stem bark on adult Wistar rat testis

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In medical practice, *Anacardium occidentale* stem bark is used for various ailment treatments. Possible histological effects of aqueous extract of *A. occidentale* stem bark administration was studied on adult Wistar rat testes. The rats (n=24), weighing 164 to 210 g were randomly assigned into three treatments (n=18) and control (n=6) groups. The rats in the treated groups II, III, IV received 20, 40 and 60 mg/kg aqueous extract of *A. occidentale* stem bark orally via orogastric tube for twenty-one days, while the control group I received only equal volume of phosphate buffer saline (PBS). The rats were fed with growers' mash and given water liberally. Cage side examination was done daily to observe overt signs of toxicity (salivation, lacrimation, yellowing of fur e.t.c). Twenty-four hours after the expiration of the extract administration, rats were sacrificed; the testes were excised and fixed in Bouin’s fluid for routine hematoxylin and eosin histological study. The histological findings of testes sections indicated that there were no cyto-architectural distortions and no evidence of apoptotic bodies in the treated groups as compared to the control group. These findings suggest that aqueous extract of *A. occidentale* does not have effect on the adult Wistar rat testis.

**Key words:** *Anacardium occidentale*, cyto-architectural, non-toxic, adult male Wistar rat.

INTRODUCTION

Plants and animals are interrelated in their environment. Plants serve as sources of food and medicinal herbs for Man and other animals in their environment. Thousands of years ago, Man has depended on plants in order to provide solutions to the myriad of health problems plaguing Him (Oliver, 1960).

In the last 20 years, the interest in medicinal plants has increased together with the number of investigations into their biological effects on human beings and animals (Veiga et al., 2005). Although, poisonous plants are ubiquitous, herbal medicine is used by up to 80% of the population in the developing countries (Jaouad et al., 2004).

Medicinal plants are believed to be an important source of new chemical substances with potential therapeutic effects (Farnsworth, 1989; Eisner, 1990).

The cashew (*Anacardium occidentale*) is a tree in the family of the flowering plant Anacardiaceae. The family contains 73 genera and about 600 species (Correa, 1978). *Anacardium* contains 8 species, native to tropical America, of which the cashew is by far the most important economically (Morton, 1987). It is a multipurpose tree of the Amazon that grows up to 15 m high (Morton, 1987; de Souza et al., 1992).

Cashew gum is used in pharmaceuticals and as substitute for gum Arabic (Patro and Behera, 1979). Extracts from roots, stems and fruits of *A. occidentale* L., (Sokeng et al., 2001) have been used by the Cameroonian and other countries’ folk medicine. In the traditional Nigerian and Brazilian pharmacopoeia, stem-bark of *A. occidentale* L. is known for its anti-inflammatory effects (Mota et al., 1985; Chen and Chung,
2000; Ojewole, 2004). The leaves and/or the bark is also used in Brazil for eczema, psoriasis, scrofula, dyspepsia, genital problems, and venereal diseases, as well as for impotence, bronchitis, cough, intestinal colic, leishmaniasis, and syphilis-related skin disorders (Franca et al., 1993). A strong antioxidant capacity was also observed against hepatocarcinogenesis induced by aflatoxin B1 in Wistar mice (Premalatha and Sachdanandam, 1999).

Preclinical studies realized with metabolites isolated from the bark of this tree species demonstrated antipyretic action for anacardic acid (Eichbaum, 1988). Polasa and Rukmini (1987) realized mutagenic tests with vegetable oil derived from cashew nut, which presented mutagenicity with or without the activation of the S9 fraction in Salmonella typhimurium. However, Melo-Cavalcante et al. (2003) and Venkatachalam and Sathe (2006) demonstrated that tannic acid, a compound present in the cashew, presented an antimutagenic effect on the Salmonella typhimurium TA 98 lineage. The bark and leaves of cashew are a rich source of tannins, a group of plant chemicals with documented biological activity (Lauren, 1999). Several clinical studies have shown that these chemicals curb the darkening effect of aging by inhibiting tyrosinase activity, and that they are toxic to certain cancer cells (Kubo et al., 1994).

Ofusori et al. (2008) reported that A. occidentale do not have a toxic effect on the brain and kidney of Swiss albino mice. Tedong et al. (2007) reported that extract amount less than 6 g/kg is not toxic for mice and high doses may have side effects.

There are no reports in the literature on the effect of aqueous extract of A. occidentale (cashew) stem bark on the testis of adult Wistar rats. The present study therefore was undertaken to evaluate the effect of this plant extract on the testis of the Wistar rats.

MATERIALS AND METHODS

The study was approved by the Research Ethical Committee of the Faculty of Basic Medical Sciences, College of Health Sciences University of Ilorin, Ilorin, Nigeria.

Plant material

Samples of A. occidentale were collected within Obafemi Awolowo University Senior Staff Quarters (Road 18), Ile-Ife in Osun State. Botanical identification of the plant was done at the herbarium section in the Department of Botany University of Ilorin, Ilorin.

Extract preparation

Fresh stem bark of A. occidentale was air dried at room temperature and pulverized with pestle and mortar. 120 g of the coarse powder was macerated with 1000 mls distilled water and the mixture was left for 48 h, mixture was filtered and filtrate was concentrated in a vacuum at 20°C (Ofusori et al., 2008) to yield the dark brown extract (paste). The extract weighed 21 g after the vacuum concentration using Gallenkamp (FA2104A, England). The solid extract was stored in a refrigerator until use (-20°C).

Experimental animals

Twenty-four adult male Wistar rats purchased from the animal house of the Department of Biochemistry, University of Ilorin with their weight ranging between 164 to 210 g were bred in the animal house of the Department of Anatomy, University of Ilorin. The rats were housed and maintained under standard conditions (12 h light/dark cycles). Food (pellet from Bendel Feeds, Yoruba Road, Ilorin) and distilled water was given ad libitum. The animals were care for in accordance with the recommendations provided in the "Guide for the Care and Use of Laboratory Animals" prepared by the National Academy of Sciences (NIH, 1985).

Grouping of animals

The animals were randomly divided into four groups of six animals each, (a control (group I), and 3 experimental groups II, III, IV). Body weights of the animals were obtained using a weighing scale (Gallenkamp FA2104A, England).

Administration of extract

Extract was administered orally to the rats by carefully inserting the syringe with the needle affix into the oral cavity of the rats. Group I serves as the control group and received phosphate buffered saline while Group II, III, IV received 20, 40 and 60 mg/kg extract, respectively for 21 days. Cage side examination were performed daily to detect overt signs of toxicity (salivation, lacrimation, chewing jaw movements, ptosis, squinting, writhing, convulsion, tremors, yellowing of fur, loss of hair, stress, erection of fur, vocalization and exophthalmia, behavioural abnormalities and dead rats) (Ratnasooriya et al., 2003). After 24 h of the last administration, the animals were sacrificed by cervical dislocation and the testes was excised and fixed in Bouin's fluid for histological studies.

Histological techniques

Paraffin tissue processing

Excised testes were fixed by immersion for the routine histological studies using Bouin’s fluid for 18 h, dehydrated through series of graded alcohol, cleared in xylene, infiltrated and embedded in molten paraffin wax. Tissue blocks were sectioned at 6µm thickness deparaffinized and stained with Haematoxylin and Eosin. The sections were examined with the light microscope and photomicrographs of the sections were taken for further analysis.

RESULTS

Physical and behavioural observation

From observation made, there was no difference in behavioural changes noticed between the control group
and experimental groups. There was no mortality, stress or abnormal behavioural changes observed within the treated groups throughout the administration periods. Overt signs of toxicity such as lacrimation, squinting and, exophthalmia etc, were not expressed by the rats throughout period of study.

**Histological Observations**

The histological observation of the testes showed no distortion in cell architectures of the treated groups. The seminiferous epithelium (SE), interstitial tissue (IT) and CVT – convoluted tubules – CVT of the treated groups showed no difference from that of the control (Figure 1a to d).

**DISCUSSION**

Effects of *A. occidentale* stem bark on the testis were investigated to highlights the possible histological implications that could result following its consumption. The study reveals that aqueous extract of cashew stem bark has no toxic effect on the testis cyto-architecture. The pharmacological mechanism behind this result may be traced to some phytochemical contents of *A. occidentale*. The stem bark of cashew is adopted in Africa as antimalaria therapy. Ngadjui et al. (2002) noted in his work on ‘antimalarial agents from some Cameroonian plants’ that, abiatane diterpenoids, quinines, triterpenoids and flavonoids are the principal phytochemical contents of some of the plants screened. Although some of these phytochemicals may not have
been successfully isolated in *A. occidentale* as literature is scanty in this area, it is believed that some of its phytochemicals may be synonymous to that found in some other plants such as *Croton zambesicus* also used traditionally in Nigeria as antimalaria therapy. Flavonoids, alkaloids and tannins have been investigated by Abo et al. (1999) and Nweze et al. (2004) to possess antimicrobial effects in various studies using plant extracts. However, more research is required to determine the role of tannins and other phytochemicals in *A. occidentale*.

However, the toxicity of *A. occidentale* L. has not been extensively studied. Paris et al. (1977) observed no toxic effects in mice treated with aqueous extracts of *A. occidentale* at doses of up to 2 g/kg by oral route while the same extract taken by *i.p.* route showed that LD<sub>50</sub> is 0.250 g/kg.

In a study, acute, 30-day sub acute toxicity and genotoxicity assays were carried out. The crude extract did not produce toxic symptoms in rats in doses up to 2000 mg/kg (Koran et al., 2007). Tedong et al. (2007) reported that extract amount less than 6 g/kg is not toxic for mice and high doses may have side effects.

Malini and Vanithakumari (1990) indicated that chronic administration of β-sitosterol (lipids isolated from *A. occidentale* leaf extract) subcutaneously to rats for 60 days was well tolerated and there was no clear evidence of any lesions either in the liver or in the kidney. Although the fruits of *A. occidentale* are commonly eaten, leaves and barks have been shown to display a wide spectrum of biological and pharmacological activities, which provide experimental support for empirical ethnopharmacologcal use of this plant in folk medicine. *Anacardium* has been reported to have anti-inflammatory actions (Mota et al., 1985; Ojewole, 2004), antimicrobial (Laurens, 1999), hypoglycaemic and anti diabetic (Kamthouing et al., 1998; Ojewole, 2004; Tedong, 2006) and molluscicidal (Mendes et al., 1990) activity. The folkloric used of *A. occidentale* may be validated by this study since the therapeutic dose often used is higher to the dose exhibiting non-toxicity in the study.

In conclusion, the functions of the testis do not influenced from cashew stem bark juice doses. We recommend that the study may be the baseline for *A. occidentale* investigations about antimicrobial and anti-inflammatory effects on experimental testis studies like orchitis.

REFERENCES


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